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The Zebrafish Neurophenome Database (ZND): A Dynamic Open-Access Resource for Zebrafish Neurophenotypic Data

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Abstract

Zebrafish (*Danio rerio*) are widely used in neuroscience research, where their utility as a model organism is rapidly expanding. Low cost, ease of experimental manipulations, and sufficient behavioral complexity make zebrafish a valuable tool for high-throughput studies in biomedicine. To complement the available repositories for zebrafish genetic information, there is a growing need for the collection of zebrafish neurobehavioral and neurological phenotypes. For this, we are establishing the Zebrafish Neurophenome Database (ZND; www.tulane.edu/~znindex/search) as a new dynamic online open-access data repository for behavioral and related physiological data. ZND, currently focusing on adult zebrafish, combines zebrafish neurophenotypic data with a simple, easily searchable user interface, which allow scientists to view and compare results obtained by other laboratories, using various treatments in different testing paradigms. As a developing community effort, ZND is expected to foster innovative research using zebrafish by federating the growing body of zebrafish neurophenotypic data.

Introduction

ZEBRAFISH (*DANIO RERIO*) ARE WIDELY USED IN NEUROSCIENCE, and their utility in behavioral research is rapidly expanding.¹⁻⁸ Although larval zebrafish have historically been popular in biomedicine,⁹⁻¹² adult zebrafish represent a novel animal model with substantial complexity and homology to higher vertebrates.^{13,14} The fully sequenced genome¹⁵⁻¹⁷ and complex behavior of zebrafish allow researchers to identify genetic and environmental factors involved in the regulation of various brain functions in this model organism.^{2,18-25}

In the age of information technology, scientists are increasingly turning to bioinformatics to find new ways of solving biological problems.²⁶⁻³² Collecting massive amounts of data in publicly available online locations has produced multiple scientific databases that make scientific information available to more scientists in more places.³³⁻⁴⁵ This "database revolution"^{47,48} has allowed researchers to apply *in silico* approaches (in addition to *in vivo* and *in vitro* experiments⁴⁹⁻⁵⁴) to analyze large amounts of data rapidly, as well as to deconstruct and re-integrate highly specific aspects of their data,

leading to a greater collective understanding of neurobiological phenomena.

Given the importance of zebrafish for modeling complex brain disorders,^{21,55-58} mounting neurobehavioral evidence requires a data repository available to scientists involved in zebrafish research. Several existing popular online databases that use other species, such as the Mouse Phenome Database (MPD), Mouse Genome Informatics (MGI), and PhenoGen,^{33,59,60} facilitated and inspired this project. These databases are excellent examples of powerful and useful online resources that foster innovative research by collecting existing behavioral and physiological data, allowing scientists to identify candidate genes, biological pathways and potential therapeutic targets.

With the recent surge of bioinformatics-based approaches, the scientific community⁶¹⁻⁶⁴ recognizes the importance of further data dissemination and sharing. The widely used Zebrafish Information Network (ZFIN) is the main online database of zebrafish biological data,⁶⁵⁻⁶⁷ comprehensively covering genomic, proteomics, and developmental data.⁶⁸ In order to cover neurophenotypes in zebrafish, we have established the Zebrafish Neurophenome Database (ZND;

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www.tulane.edu/~znpindex/search) which currently contains adult zebrafish neurophenotypes, but will eventually include larval zebrafish models as well. ZND is a dynamic database supported by the Tulane University Innovative Learning Center (ILC) as part of the Zebrafish Neurophenome Project led by the Zebrafish Neuroscience Research Consortium (ZNRC), an international network of collaborating laboratories dedicated to zebrafish research. As an ongoing, community-driven effort,⁶⁹ this database is an online open-source repository for behavioral and related physiological data, reflecting the current state of knowledge on zebrafish neurophenotypes. The utility of this project greatly depends on contributions from the zebrafish research community,⁶⁹ and the goal of this paper is to introduce the ZND concept to a wide audience of zebrafish investigators.

Content of the ZND

ZND is designed to consolidate zebrafish information to provide a comprehensive open access database of their behavioral and neurological phenotypes. The database contains details of experimental manipulations and the corresponding responses across all major behavioral domains and tests/paradigms (Table 1), enabling a better characterization and interpretation of zebrafish phenotypes. ZND also reports negative findings (e.g., ineffective drug doses or inactive stressors), which gives researchers complete information to make more informed decisions regarding optimal experimental design of their zebrafish studies.

T1 ▶

F1 ▶

The information flow between ZND, its contributors, and users is shown in Figure 1. Using the input of the growing zebrafish neuroscience community, ZND covers various forms of scientific communication including scholarly journal publications, books and book chapters, conference abstracts, patents, doctoral dissertations, theses, project reports, government documents, web-sites, posters, personal communications, and unpublished observations. ZND also allows users to submit their own data, which are curated (to assure compatibility) by its team and then added to the database. For this, a detailed Excel template and online submission forms are provided on the ZND web-site within the "Submit"

toolbox. Users can review their uploaded data, verify its accuracy, and contact the ZND team to make necessary corrections (Fig. 1). Since the original publications may occasionally omit important methodological details (e.g., pre-treatment duration or testing intervals) or results, the ZND team will contact the principal investigators for clarification, in order to correctly update this information to the best of our knowledge. Collectively, this is expected to facilitate the comprehensive and updated coverage of zebrafish neurophenotypic data, making it accessible to researchers in this field.

Organization and Structure of the ZND

In its present form, ZND is maintained on a Tulane University server running an industry-standard Linux-Apache-MySQL/PHP application stack (LAMP) as a MySQL database. The database is hosted on a secure up-to-date professional-grade web server named Pulse, with automatic nightly back-up and a hurricane disaster recovery plan. The ZND data repository is managed using VFront,⁷⁰ a free open source user front-end for managing data within MySQL and PostgreSQL databases. The search interface and results screens are authored, supported, and continuously developed by the ILC of Tulane Technology Services.

The summary of a typical experiment in ZND is presented in Figure 2. Each *study*, which contains detailed descriptions of the age, sex, strain, and housing conditions of the fish used, is divided into *intervals* based on the timing of the experiment (a 5-day treatment exposure has 5 intervals of 1-day each, or a single interval of 5 days).⁷¹ The intervals are further separated by *treatment*, which can be any behavioral, pharmacological, or genetic manipulation (e.g., predator stress, acute morphine exposure or mutant strain, respectively). Each treatment also contains *test* details such as *model* and its *duration* (e.g., 6-min novel tank test, 15-min light/dark box, 30-min open field test) and *test results*. For each treatment and test, all endpoints are listed in the results section, with a field for treatment effect (i.e., increased or decreased compared to controls) and a field denoting statistical significance. Specific endpoints which were not significantly modulated in a given experiment are listed as "Unaffected endpoints" on the results page, reflecting all negative results from each study in ZND.

◀ F2

To search ZND for neurobehavioral data, users can select an experiment or treatment from a drop-down list of treatments (on the database's main search page), ranging from pharmacological manipulations (e.g., acute cocaine or nicotine) to experimental stressors (e.g., light exposure or vortex stress).⁷¹ The endpoints measured for a given treatment are listed in the result output. Other search options, such as by the Principal Investigator's name, are also available in the ZND,⁷¹ and will be expanded in the future.

Potential Applications

One of the goals of the ZND is to increase our knowledge and basic understanding of zebrafish neurobiology. For example, a zebrafish researcher working with glutamatergic drugs can search ZND data for the selected 'reference' treatments (e.g., Ketamine or MK-801), and use results generated in a different laboratory, to determine the effective dose range of these compounds in adult zebrafish, and/or to compare their effects with responses observed in his own laboratory.

TABLE 1. GENERAL SIMILARITY OF SELECTED ZEBRAFISH AND RODENT BEHAVIORAL DOMAINS AND EXPERIMENTAL PARADIGMS

Rodent Phenotype/Tests	Fish Phenotype/Tests
Freezing behavior/open field ^{81,82}	Freezing behavior/open field, novel tank ¹⁹
Hyperlocomotion/open field ^{83,84}	Hyperactivity/erratic movements ^{85,86}
Thigmotaxis/open field ^{87,88}	Thigmotaxis/open field ^{89,90}
Scototaxis/light-dark box ^{91,92}	Scototaxis/light-dark box ^{93,94}
Homebase formation/open field ⁹⁵	Homebase formation/open field ⁹⁶
Novelty habituation/open field ⁹⁷	Novelty habituation/open field ⁹⁸
Social preference ^{99,100}	Social preference and shoaling ^{101,102}

See also References 8, 13, 18, 21, and 80 for review.

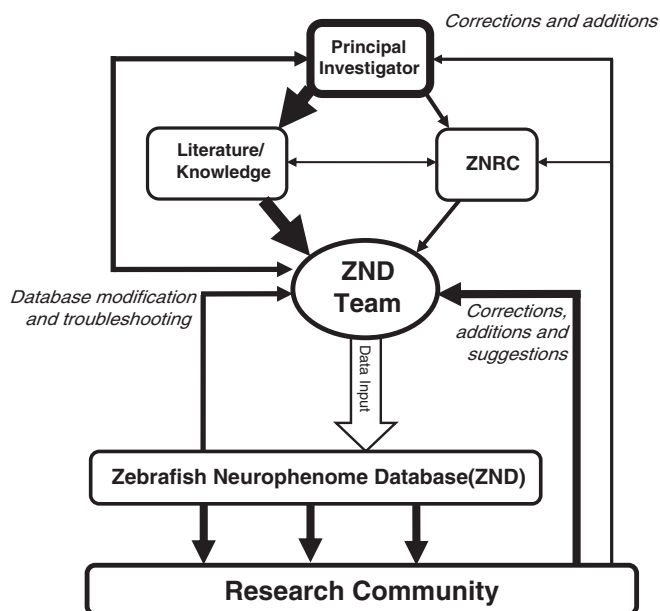


FIG. 1. A diagram summarizing the Zebrafish Neurophenome Database (ZND) development and information flow. This diagram displays the relationships between principal investigators interested in zebrafish research, the Zebrafish Neuroscience Research Consortium (ZNRG), and the ZND framework. The ZND inter-disciplinary team currently consists of zebrafish neuroscientists (Kalueff laboratory at Tulane University, New Orleans, LA, www.kaluefflab.com), computer/database team (Tulane Innovative Learning Center), and scientific literature experts (Tulane Howard-Tilton Memorial Library). In addition to data search by the ZND team, the investigators are able to correct data input into the database, ensuring its reliability and accuracy. ZND also regularly networks with zebrafish investigators, inviting them to review and update data currently available in the database, as well as submit their most recent findings.

◀AU1

Scientists can also use the concentrations and resulting effects of those treatments to determine the dose range for other drugs not previously tested in zebrafish, but whose activity dose ranges may be extrapolated based on potency relative to other ligands that are already in the database. ZND also improves our ability to critically evaluate the available zebrafish data. For example, peer-reviewers of journal articles, grant applications, or protocols submitted to the Institutional Animal Use Committees (IACUC), who are not experts in zebrafish neuroscience, may benefit from using ZND to evaluate the novelty of proposed research, feasibility of approaches and the efficacy of proposed treatments.

ZND also assists interdisciplinary research across different domains of knowledge. For example, a researcher searching for environmental contaminants in adult zebrafish can use ZND to determine which compounds are already known to be toxic to zebrafish, thereby maximizing time and re-

sources for prospective studies and establishing effective dose ranges. Statistical tools and raw data, albeit currently not part of ZND, will eventually be added to the database (similar to MPD, MGI, and PhenoGen databases) to assist in such analyses.

Furthermore, ZND allows researchers to plan experiments better, saving time and resources in accordance with the 3R's principles of research.^{72,73} For example, the use of ZND can reduce the number of animals involved in the planned study by allowing scientists to determine effective dose ranges and their most sensitive endpoints, based on prior research performed by other laboratories. The database also helps to refine the existing techniques used to study zebrafish behavior by displaying data collected in the most common behavioral testing paradigms, thereby facilitating further analysis by identifying the best models for testing a particular treatment in question. Such *in silico* experimentation also complements and optimizes animal research by enabling re-analysis and meta-analysis of existing data without the need to repeat actual testing in live animals. The possibility of rapid analysis of zebrafish phenotypic data improves data-mining ability and allows for the efficient assessment of complex behavioral domains.

Importantly, ZND can also be utilized as a complement to literature searches. For example, ZND collects positive and negative findings from published and unpublished data in multiple formats, including book chapters, conference abstracts, and personal communications (many of which are not covered by the PubMed and other traditional tools for scientific literature search). Using ZND, researchers can now collect and analyze published and unpublished data in a shorter period of time, uncovering novel relationships and improving our understanding of the zebrafish models. Likewise, investigators working with other popular fish species (such as goldfish or medaka fish), may also benefit from ZND, using it, for example, to compare their own results with those generated in zebrafish models.

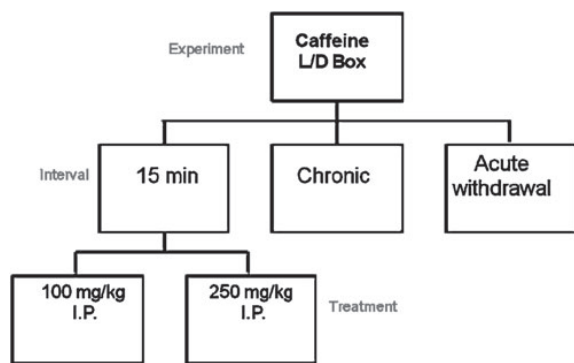


FIG. 2. Diagram illustrating parent-child relations within different components in the Zebrafish Neurophenome Database (ZND).

As already mentioned, the importance of sharing and dissemination of scientific knowledge is increasingly recognized by the scientific community,^{61–64} federal funding agencies,^{74,75} and national legislation in various countries. Other funding agencies and leading academic journals also require that authors of peer-reviewed papers share their raw data, statistical methods, and software, necessary to interpret or reproduce their published research. The zebrafish research community fully supports the need for data sharing and dissemination.^{65–67} In line with this, ZND offers an opportunity for zebrafish laboratories to *upload* and *store* their data, serving as a free data repository for investigators working in the field of zebrafish neuroscience. Finally, in addition to promoting zebrafish research, ZND can also be used in biomedical education and teaching, where the potential of this model organism is becoming widely recognized.^{65,76–79} For example, high-school and college educators can use ZND to identify effective treatments and experimental models to be used for classroom demonstrations and independent research projects of their students.

Overall, ZND is designed as a dynamic framework that is constantly enhanced and modified to incorporate novel findings in the field in close contact with zebrafish laboratories and the neuroscience research community (Fig. 1). The ZND team will continue to expand the database to incorporate genetic and physiological evidence, especially genomic, proteomics, and pathway data in order to more fully characterize adult zebrafish behavior and neurophysiology. With the growing international network of laboratories involved in zebrafish research, ZNRC and other members of the zebrafish research community⁶⁹ will continue to provide necessary expert support to ZND, further increasing the participation of established and newer groups in sharing their zebrafish data through ZND.

Conclusion

In summary, ZND is expected to continue to enhance zebrafish neurophenotyping by providing scientists with an open-access comprehensive resource of relevant data generated in this species. This innovative, dynamic online database offers a timely tool for researchers studying zebrafish behavior and neurobiology. Continuing the current trends of data sharing and dissemination, it increases visibility of zebrafish research, and encourages collaboration between established and new laboratories worldwide. Providing the researchers with effective new resource for data-mining and analysis, ZND shows how compact *specialized* neurobiological databases not only address specific biomedical problems, but can also advance out understanding of complex biological phenomena.

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References

- Gerlai R, Lee V, Blaser R. Effects of acute and chronic ethanol exposure on the behavior of adult zebrafish (*Danio rerio*). *Pharmacol Biochem Behav* 2006;85:752–761.
- Zon LI. Zebrafish: A new model for human disease. *Genome Res* 1999;9:99–100.
- Cachat J, Stewart A, Utterback E, Hart P, Gaikwad S, Wong K, et al. Three-dimensional neurophenotyping of adult zebrafish behavior. *PLoS One* 2011;6:e17597.
- Maximino C, da Silva AW, Gouveia A, Jr., Herculano AM. Pharmacological analysis of zebrafish (*Danio rerio*) scototaxis. *Prog Neuropsychopharmacol Biol Psychiat* 2011;35:624–631.
- Ahmed O, Seguin D, Gerlai R. An automated predator avoidance task in zebrafish. *Behav Brain Res* 2011;216:166–171.
- Sackerman J, Donegan JJ, Cunningham CS, Nguyen NN, Lawless K, Long A, et al. Zebrafish behavior in novel environments: Effects of acute exposure to anxiolytic compounds and choice of *Danio rerio* line. *Int J Comp Psychol* 2010;23:43–61.
- Piato AL, Capiotti KM, Tamborski AR, Oses JP, Barcellos LJ, Bogo MR, et al. Unpredictable chronic stress model in zebrafish (*Danio rerio*): Behavioral and physiological responses. *Prog Neuropsychopharmacol Biol Psychiatry* 2011;35:561–567.
- Miklosi A, Andrew RJ. The zebrafish as a model for behavioral studies. *Zebrafish* 2006;3:227–234.
- Lockwood B, Bjerke S, Kobayashi K, Guo S. Acute effects of alcohol on larval zebrafish: A genetic system for large-scale screening. *Pharmacol Biochem Behav* 2004;77:647–654.
- Tiedeken JA, Ramsdell JS. Embryonic exposure to domoic acid increases the susceptibility of zebrafish larvae to the chemical convulsant pentylenetetrazole. *Environ Health Perspect* 2007;115:1547–1552.
- Burgess HA, Granato M. Sensorimotor gating in larval zebrafish. *J Neurosci* 2007;27:4984–4994.
- Jin M, Zhang X, Wang L, Huang C, Zhang Y, Zhao M. Developmental toxicity of bifenthrin in embryo-larval stages of zebrafish. *Aquat Toxicol* 2009;95:347–354.
- Lieschke GJ, Currie PD. Animal models of human disease: Zebrafish swim into view. *Nat Rev Genet* 2007;8:353–367.
- Brittijn SA, Duivesteyn SJ, Belmamoune M, Bertens LF, Bitter W, de Bruijn JD, et al. Zebrafish development and regeneration: New tools for biomedical research. *Int J Dev Biol* 2009;53:835–850.
- Ekker SC, Stemple DL, Clark M, Chien CB, Rasooly RS, Javois LC. Zebrafish genome project: Bringing new biology to the vertebrate genome field. *Zebrafish* 2007;4:239–251.
- Jekosch K. The zebrafish genome project: Sequence analysis and annotation. *Methods Cell Biol* 2004;77:225–239.
- Woods IG, Kelly PD, Chu F, Ngo-Hazelett P, Yan YL, Huang H, et al. A comparative map of the zebrafish genome. *Genome Res* 2000;10:1903–1914.
- Cachat JM, Canavello PR, Elegante MF, Bartels BK, Elkhayat SI, Hart PC, et al. Modeling stress and anxiety in zebrafish. In: *Zebrafish Models in Neurobehavioral Research*. Kalueff AV, Cachat J, (eds), pp. 73–88. Humana Press, New York, 2010.

19. Egan RJ, Bergner CL, Hart PC, Cachat JM, Canavello PR, Elegante MF, et al. Understanding behavioral and physiological phenotypes of stress and anxiety in zebrafish. *Behav Brain Res* 2009;205:38–44.
20. Levin ED, Limpuangthip J, Rachakonda T, Peterson M. Timing of nicotine effects on learning in zebrafish. *Psychopharmacology (Berl)* 2006;184:547–552.
21. Maximino C, de Brito TM, da Silva Batista AW, Herculano AM, Morato S, Gouveia A, Jr. Measuring anxiety in zebrafish: A critical review. *Behav Brain Res* 2010;214:157–171.
22. Seibt KJ, Oliveira Rda L, Zimmermann FF, Capiotti KM, Bogo MR, Ghisleni G, et al. Antipsychotic drugs prevent the motor hyperactivity induced by psychotomimetic MK-801 in zebrafish (*Danio rerio*). *Behav Brain Res* 2010;214:417–422.
23. To TT, Hahner S, Nica G, Rohr KB, Hammerschmidt M, Winkler C, et al. Pituitary-interrenal interaction in zebrafish interrenal organ development. *Mol Endocrinol* 2007;21:472–485.
24. von Krogh K. Environmental enrichment and its effects on telencephalic neurogenesis and behaviour in isolated adult zebrafish, *Danio rerio*. Oslo: University of Oslo; 2007.
25. Xi Y, Ryan J, Noble S, Yu M, Yilbas AE, Ekker M. Impaired dopaminergic neuron development and locomotor function in zebrafish with loss of pink1 function. *Eur J Neurosci* 2010;31:623–633.
26. Stewart A, Gaikwad S, Hart P, Kyzar E, Roth A, Kalueff AV. Experimental models for anxiolytic drug discovery in the era of omes and omics. *Expert Opin Drug Discov* 2011;1–15.
- AU2 ▶ 27. Lein ES, Hawrylycz MJ, Ao N, Ayres M, Bensinger A, Bernard A, et al. Genome-wide atlas of gene expression in the adult mouse brain. *Nature* 2007;445:168–176.
28. Liu P, Vikis H, Lu Y, Wang D, You M. Large-scale in silico mapping of complex quantitative traits in inbred mice. *PLoS One* 2007;2:e651.
29. Grupe A, Germer S, Usuka J, Aud D, Belknap JK, Klein RF, et al. *In silico* mapping of complex disease-related traits in mice. *Science* 2001;292:1915–1918.
30. Ditzen C, Varadarajulu J, Czibere L, Gonik M, Targosz BS, Hamsch B, et al. Proteomic-based genotyping in a mouse model of trait anxiety exposes disease-relevant pathways. *Mol Psychiat* 2010;15:702–711.
31. Stone N, Pangilinan F, Molloy AM, Shane B, Scott JM, Ueland PM, et al. Bioinformatic and genetic association analysis of microRNA target sites in one-carbon metabolism genes. *PLoS One* 2011;6:e21851.
32. Gerlai R. Phenomics: Fiction or the future? *Trends Neurosci* 2002;25:506–509.
33. Bhave SV, Hornbaker C, Phang TL, Saba L, Lapadat R, Kechris K, et al. The PhenoGen informatics website: Tools for analyses of complex traits. *BMC Genet* 2007;8:59.
34. Colorado-Denver Uo. PhenoGen Informatics. Denver, Colorado USA, 2011.
35. D'Eustachio P. Reactome knowledgebase of human biological pathways and processes. *Methods Mol Biol* 2011;694:49–61.
36. Croft D, O'Kelly G, Wu G, Haw R, Gillespie M, Matthews L, et al. Reactome: A database of reactions, pathways, and biological processes. *Nucleic Acids Res* 2011;39:D691–697.
- AU3 ▶ 37. Shaw DR. Searching the Mouse Genome Informatics (MGI) resources for information on mouse biology from genotype to phenotype. *Curr Protoc Bioinformatics* 2009;Chapter 1:Unit1 7.
38. Laboratory TJ. Mouse Genome Informatics. Bar Harbor, Maine USA, 2011.
39. Thorisson GA, Smith AV, Krishnan L, Stein LD. The International HapMap Project Web site. *Genome Res* 2005;15:1592–1593.
40. Consortium TIH. The International HapMap Project. ◀ AU4
41. Hamosh A, Scott AF, Amberger JS, Bocchini CA, McKusick VA. Online Mendelian Inheritance in Man (OMIM), a knowledge base of human genes and genetic disorders. *Nucleic Acids Res* 2005;33:D514–517.
42. University JH. Online Mendelian Inheritance in Man (OMIM). ◀ AU5
43. Nicholas FW. Online Mendelian Inheritance in Animals (OMIA): A comparative knowledgebase of genetic disorders and other familial traits in non-laboratory animals. *Nucleic Acids Res* 2003;31:275–277.
44. Nicholas FW. Online Mendelian Inheritance in Animals (OMIA). ◀ AU6
45. Kahraman A, Avramov A, Nashev LG, Popov D, Ternes R, Pohlenz HD, et al. PhenomicDB: A multi-species genotype/phenotype database for comparative phenomics. *Bioinformatics* 2005;21:418–420.
46. This reference has been deleted. ◀ AU7
47. Hahn U, Wermter J, Blaszczak R, Horn PA. Text mining: Powering the database revolution. *Nature* 2007;448:130.
48. Gollery M. Your database is obsolete: The promise of contextual bioinformatics. *Bioinformatics* 2007;1:356.
49. Hoffman PL, Bennett B, Saba LM, Bhave SV, Carosone-Link PJ, Hornbaker CK, et al. Using the Phenogen website for 'in silico' analysis of morphine-induced analgesia: Identifying candidate genes. *Addict Biol* 2011;16:393–404.
50. Elkins KM. An *in silico* DNA cloning experiment for the biochemistry laboratory. *Biochem Mol Biol Educ* 2011;39:211–215.
51. Takano M, Terada TP, Sasai M. Unidirectional Brownian motion observed in an *in silico* single molecule experiment of an actomyosin motor. *Proc Natl Acad Sci USA* 2010;107:7769–7774.
52. Chan CH, Mitchison HM, Pearce DA. Transcript and *in silico* analysis of CLN3 in juvenile neuronal ceroid lipofuscinosis and associated mouse models. *Hum Mol Genet* 2008;17:3332–3339.
53. Naumova EN, Gorski J, Naumov YN. Simulation studies for a multistage dynamic process of immune memory response to influenza: Experiment *in silico*. *Ann Zool Fennici* 2008;45:369–384.
54. O'Connor KA, Roth BL. Screening the receptorome for plant-based psychoactive compounds. *Life Sci* 2005;78:506–511.
55. Kokel D, Peterson RT. Chemobehavioural phenomics and behaviour-based psychiatric drug discovery in the zebrafish. *Brief Funct Genomic Proteomic* 2008;7:483–490.
56. Kari G, Rodeck U, Dicker AP. Zebrafish: An emerging model system for human disease and drug discovery. *Clin Pharmacol Ther* 2007;82:70–80.
57. Tomasiewicz HG, Flaherty DB, Soria JP, Wood JG. Transgenic zebrafish model of neurodegeneration. *J Neurosci Res* 2002;70:734–745.
58. Linker A, Stewart A, Gaikwad S, Cachat J, Elegante M, Kalueff AV, et al. Assessing the maximal predictive validity for neuropharmacological anxiety screening assays using zebrafish. In: *Zebrafish Neurobehavioral Protocols*. Kalueff AV, Cachat J, (eds). Humana Press, New York, 2010.

59. Bennett B, Saba LM, Hornbaker CK, Kechris KJ, Hoffman P, Tabakoff B. Genetical genomic analysis of complex phenotypes using the PhenoGen website. *Behav Genet* 2011;41:625–628.
60. Hoffman PL, Bennett B, Saba LM, Bhave SV, Carosone-Link PJ, Hornbaker CK, et al. Using the Phenogen website for ‘in silico’ analysis of morphine-induced analgesia: Identifying candidate genes. *Addict Biol* 2011;16:393–404.
61. Gardner D, Toga AW, Ascoli GA, Beatty JT, Brinkley JF, Dale AM, et al. Towards effective and rewarding data sharing. *Neuroinformatics* 2003;1:289–295.
62. Chervitz SA, Deutsch EW, Field D, Parkinson H, Quackenbush J, Rocca-Serra P, et al. Data standards for Omics data: The basis of data sharing and reuse. *Methods Mol Biol* 2011;719:31–69.
63. Field D, Sansone SA, Collis A, Booth T, Dukes P, Gregurick SK, et al. Megascience. ‘Omics data sharing. *Science* 2009;326:234–236.
64. Arzberger P, Schroeder P, Beaulieu A, Bowker G, Casey K, Laaksonen L, et al. Science and government. An international framework to promote access to data. *Science* 2004;303:1777–1778.
65. Sprague J, Bayraktaroglu L, Bradford Y, Conlin T, Dunn N, Fashena D, et al. The Zebrafish Information Network: The zebrafish model organism database provides expanded support for genotypes and phenotypes. *Nucleic Acids Res* 2008;36:D768–772.
66. Sprague J, Clements D, Conlin T, Edwards P, Frazer K, Schaper K, et al. The Zebrafish Information Network (ZFIN): The zebrafish model organism database. *Nucleic Acids Res* 2003;31:241–243.
67. Westerfield M, Doerry E, Kirkpatrick AE, Douglas SA. Zebrafish informatics and the ZFIN database. *Methods Cell Biol* 1999;60:339–355.
68. Sprague J, Doerry E, Douglas S, Westerfield M. The Zebrafish Information Network (ZFIN): A resource for genetic, genomic and developmental research. *Nucleic Acids Res* 2001;29:87–90.
- AU8 ▶ 69. authors) SoSm: Statement of Support. 2011.
- AU9 ▶ 70. Verona M. VFront. Provincia di Cagliari, Italy, 2007.
71. Zapolsky I, Kyzar E, Green J, Gaikwad S, Pham M, Chanin S, et al. Utilizing the Zebrafish Neurophenome Project (ZNP) Database for analyses of complex neurophenotypes in zebrafish models. In: *Zebrafish Protocols for Neurobehavioral Research*. Kalueff A, Stewart A, (eds). Humana Press, New York, 2011.
72. Russell WMS, Burch RL *The Principles of Humane Experimental Technique*. London: Methuen; 1959.
73. Flecknell P. Replacement, reduction and refinement. *ALTEX* 2002;19:73–78.
74. Health Nio. NIH Data Sharing Policy. 2003.
- AU10 ▶ 75. Foundation NS. Dissemination and Sharing of Research Results.
76. Shuda J, Kearns-Sixsmith D. Outreach: Empowering students and teachers to fish outside the box. *Zebrafish* 2009;6:133–138.
77. Bilotta J, Saszik S, DeLorenzo AS, Hardesty HR. Establishing and maintaining a low-cost zebrafish breeding and behavioral research facility. *Behav Res Methods Instrum Comput* 1999;31:178–184.
78. McKeown KA, Downes GB, Hutson LD. Modular laboratory exercises to analyze the development of zebrafish motor behavior. *Zebrafish* 2009;6:179–185.
79. Hutson LD, Liang JO. Making an impact: Zebrafish in education. *Zebrafish* 2009;6:119–120.
80. Norton W, Bally-Cuif L. Adult zebrafish as a model organism for behavioural genetics. *BMC Neurosci* 2010;11:90.
81. Conti LH, Maciver CR, Ferkany JW, Abreu ME. Footshock-induced freezing behavior in rats as a model for assessing anxiolytics. *Psychopharmacology (Berl)* 1990;102:492–497.
82. Fanselow MS. Conditioned and unconditional components of post-shock freezing. *Pavlov J Biol Sci* 1980;15:177–182.
83. Gould TD, O’Donnell KC, Picchini AM, Manji HK. Strain differences in lithium attenuation of d-amphetamine-induced hyperlocomotion: A mouse model for the genetics of clinical response to lithium. *Neuropsychopharmacology* 2007;32:1321–1333.
84. Irifune M, Sato T, Nishikawa T, Masuyama T, Nomoto M, Fukuda T, et al. Hyperlocomotion during recovery from isoflurane anesthesia is associated with increased dopamine turnover in the nucleus accumbens and striatum in mice. *Anesthesiology* 1997;86:464–475.
85. Darland T, Dowling JE. Behavioral screening for cocaine sensitivity in mutagenized zebrafish. *Proc Natl Acad Sci USA* 2001;98:11691–11696.
86. Williams LR, Wong K, Stewart A, Suci C, Gaikwad S, Wu N, et al. Behavioral and physiological effects of RDX on adult zebrafish. *Comp Biochem Physiol C Toxicol Pharmacol* 2012;155:33–38.
87. Lamprea MR, Cardenas FP, Setem J, Morato S. Thigmotactic responses in an open-field. *Braz J Med Biol Res* 2008;41:135–140.
88. Choleris E, Thomas AW, Kavaliers M, Prato FS. A detailed ethological analysis of the mouse open field test: Effects of diazepam, chlordiazepoxide and an extremely low frequency pulsed magnetic field. *Neurosci Biobehav Rev* 2001;25:235–260.
89. Blaser RE, Chadwick L, McGinnis GC. Behavioral measures of anxiety in zebrafish (*Danio rerio*). *Behav Brain Res* 2010;208:56–62.
90. Grossman L, Utterback E, Stewart A, Gaikwad S, Chung KM, Suci C, et al. Characterization of behavioral and endocrine effects of LSD on zebrafish. *Behav Brain Res* 2010;214:277–284.
91. Belzung C, Griebel G. Measuring normal and pathological anxiety-like behaviour in mice: A review. *Behav Brain Res* 2001;125:141–149.
92. Belzung C, Misslin R, Vogel E. Anxiogenic effects of a benzodiazepine receptor partial inverse agonist, RO 19-4603, in a light/dark choice situation. *Pharmacol Biochem Behav* 1990;36:593–596.
93. Maximino C, Marques de Brito T, Dias CA, Gouveia A, Jr., Morato S. Scototaxis as anxiety-like behavior in fish. *Nat Protoc* 2010;5:209–216.
94. Maximino C, de Brito TM, Colmanetti R, Pontes AA, de Castro HM, de Lacerda RI, et al. Parametric analyses of anxiety in zebrafish scototaxis. *Behav Brain Res* 2010;210:1–7.
95. Eilam D, Golani I. Home base behavior of rats (*Rattus norvegicus*) exploring a novel environment. *Behav Brain Res* 1989;34:199–211.
96. Stewart A, Cachat J, Wong K, Gaikwad S, Gilder T, DiLeo J, et al. Homebase behavior of zebrafish in novelty-based paradigms. *Behav Processes* 2010;85:198–203.
97. Rowe WB, Spreekmeester E, Meaney MJ, Quirion R, Rochford J. Reactivity to novelty in cognitively-impaired and cognitively-unimpaired aged rats and young rats. *Neuroscience* 1998;83:669–680.

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98. Wong K, Elegante M, Bartels B, Elkhayat S, Tien D, Roy S, et al. Analyzing habituation responses to novelty in zebrafish (*Danio rerio*). *Behav Brain Res* 2010;208:450–457.
99. Van Loo PL, Van de Weerd HA, Van Zutphen LF, Baumans V. Preference for social contact versus environmental enrichment in male laboratory mice. *Lab Anim* 2004;38:178–188.
100. Winslow JT. Mouse social recognition and preference. *Curr Protoc Neurosci* 2003;Chapter 8:Unit 8 16.
101. Saverino C, Gerlai R. The social zebrafish: Behavioral responses to conspecific, heterospecific, and computer animated fish. *Behav Brain Res* 2008;191:77–87.
102. Engeszer RE, Ryan MJ, Parichy DM. Learned social preference in zebrafish. *Curr Biol* 2004;14:881–884.

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